



The Effect of Shaking, CO₂ Concentration and Light Intensity on Biomass Growth of Green Microalgae *Desmodesmus communis*

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There are many factors that can affect microalgae growth. In this research, four different groups of experiments were set up in order to determine the influence of different mixing conditions, CO₂ concentration and light intensities on *Desmodesmus communis* growth. The range of CO₂ concentration in the air-CO₂ mixture was 0–16 v/v%, light intensities ranged between 100 μmol m⁻²s⁻¹ and 300 μmol m⁻²s⁻¹. The best biomass productivity and biomass yield of 0.54 g d⁻¹ and 3.53 g l⁻¹ respectively were achieved when mixing was provided by using shaker as well as gas bubbling with air-CO₂ mixture of 96:4 v/v% and light intensity of 300 μmol m⁻²s⁻¹.

Keywords: *microalgae, mixing, CO₂ concentration, light intensities, biomass growth.*

1 Introduction

Microalgae are relatively easy to cultivate, they can grow using wastewater as well as clean water and can easily obtain nutrients (Makarevičienė et al., 2011; Samori et al., 2013; Sheenan et al., 1998). Using photosynthesis, microalgae can reproduce themselves and complete all growth stages in a short time (Sheenan et al., 1998). Because of variety of high-value biological derivatives, they can produce a wide range of feedstock for biofuel production and also for many other applications, for example, microalgae are also used in cosmetics, pharmaceuticals, nutrition and food additives, aquaculture, and pollution prevention (Makarevičienė et al., 2011; Makarevičienė et al., 2012; Mata et al., 2010; Schenk et al., 2008).

Those algae cultures from which producing of high value products is possible are usually cultivated under sterile conditions with relatively well-defined external parameters. Specially built photobioreactors can provide such conditions (Scott et al., 2010). They allow cultivation of single-species microalgae culture for long term and have been successfully used for

producing large quantities of microalgal biomass (Chisti, 2007). Photobioreactors can operate in batch, fed-batch and continuous mode. Batch culture is one of the most used, and it can be characterised as a closed system, in which the volume is limited and resources are finite – cell density continuously increases until one or more of the limiting factors ends (Barsanti and Gualtieri, 2006).

Nevertheless, algae cultivation has its own challenges as well. One of the most critical links in microalgae research and application is strain selection – a need to select and grow highly productive algal strains (Han et al., 2012; Rodolfi et al., 2009). Then the energy consumption follows. Closed photobioreactors need energy for mixing the culture suspension, water pumping, gas compressing for bubbling (CO₂ transfer), harvesting/dewatering the culture biomass, and they have much embodied energy in the materials of construction (Rodolfi et al., 2009; Scott et al., 2010). One more limitation for large-scale production is the availability of the

sources of nutrients; often chemical or inorganic fertilizers are used to achieve appropriate growth rate of microalgae (Lam et al., 2013). Moreover, even though CO₂ may be available as flue gases from power or chemical plants on industrial scale, the distribution of CO₂ is problematic (Scott et al., 2010). Still in many cases, because of the benefits of algae cultivation, it is worth dealing with and overcoming these challenges.

Biomass production is remarkably affected by environmental factors (Fang et al., 2013). The ones that influence the growth the most are light intensity and light-dark cycle, temperature, and nutrients status – concentrations of carbon and nitrogen, pH (Fang et al., 2013; Li et al., 2011). It is reported that mixing and aeration are also very important (Ugwu et al., 2007). Good mixing improves biomass productivity by increasing the frequency of cell exposure to light and dark volumes of the reactor as well as by increasing mass transfer between the nutrients and cells (Kunjapur et al., 2010).

The aim of this research is to study the influence of different mixing conditions, CO₂ concentration and light intensities on algae biomass productivity in order to design an appropriate photobioreactor. To do so it is necessary to clarify all the factors which influence the results of algal cultivation as well as the parameter control possibilities. The experiments were performed in shake flasks because that is one of the easiest ways to provide multi-parameter variation in parallel experiments. This is also shown in Samori et al. work (2013) where different sets of experiments for *Desmodesmus communis* cultivation were run in batch culture in order to determine the effect of medium composition, CO₂-air mixture, and light intensity on algae growth.

In the experiments performed, the authors have also used *D. communis* that is unicellular freshwater green algae.

2 Methods

2.1 Microorganism and culture medium

A novel strain of green algae was isolated from fresh water samples collected from estuary zone of River Daugava (Riga, Latvia) in August 2008. This novel strain was identified as *D. communis* named as DCDA-3 (culture collection of Latvian Institute of Aquatic Ecology).

Each bottle contained 400 ml culture medium of BG11, which consisted of 1.5 g l⁻¹ NaNO₃, 0.04 g l⁻¹ K₂HPO₄, 0.075 g l⁻¹ MgSO₄·7H₂O, 0.006 g l⁻¹ Fe(NH₄)₃(C₆H₅O₇)₂, 0.001 g l⁻¹ EDTANa₂, 0.006 g l⁻¹ citric acid, 0.02 g l⁻¹ Na₂CO₃, 0.036 g l⁻¹ CaCl₂·2H₂O, 0.001 g l⁻¹ vitamin B1 and trace element solution – 0.000040 g l⁻¹ CoCl₂·6H₂O, 0.00286 g l⁻¹ H₃BO₃, 0.00181 g l⁻¹ MnCl₂·4H₂O, 0.000222 g l⁻¹ ZnSO₄·7H₂O, 0.00004 g l⁻¹ Na₂MoO₄·2H₂O, 0.000080 g l⁻¹ CuSO₄·5H₂O. All media was autoclaved for sterilization at 120 °C for 45 min. The

volume of the seed culture used was appropriately chosen to obtain OD of 0.5 for culture media.

2.2 Experimental setup and cultivation conditions

Four parallel groups of experiments were set up for *D. communis* cultivation under different growth conditions: 1. Cultivation with aeration under static condition; 2. Cultivation with shaking and aeration; 3. Cultivation with shaking and different light intensities; 4. Cultivation with aeration, shaking and different light intensities. Schematic diagram of the experiments setup is shown in figure 1.

D. communis were grown in 500 ml DURAN® GLS 80® laboratory bottles (height 148 mm, diameter 101 mm) filled with 400 ml of culture medium. Cultivation temperature 25 ± 1 °C was provided by inserting thermostat tubes into all bottles. Illumination provided by LED (the light/dark periods were 16/8 h), aerated by bubbling air-CO₂ mixture (100:0, 99:1, 96:4, 92:8 and 84:16 v/v%), which was continuously supplied with a flow rate of 0.5 vvm (gas volume per liquid culture volume and per hour). The culture was stirred using BioSan PSU-20i multifunctional orbital shaker. For illumination, the cases were used that were made of 3 m long LED 3528 strips 9.6 W IP33 (white/cold), with 60 LEDs per meter. Light intensity for Group I and Group II in each bottle was 200 μmol m⁻²s⁻¹ but in the bottles from Group III and IV there were different light intensities: 100, 200, and 300 μmol m⁻²s⁻¹. The appropriate light intensity was adjusted using LED dimmer and measured by a Li-250A Light Meter with a Li-190 quantum sensor (LI-COR Inc., Lincoln, Nebraska USA). All the experiments were performed in duplicates.

Experimental setup for each group:

- I) Cultivation with aeration under static conditions: cultivation was performed in bottles equipped with a gas sparger and a reflux condenser.
- II) Cultivation with shaking and aeration: shaker (160 rpm), bottles equipped with gas sparger and a reflux condenser.
- III) Cultivation with shaking and different light intensities: shaker (160 rpm), bottles.
- IV) Cultivation with aeration, shaking and different light intensities: shaker (160 rpm), bottles equipped with a gas sparger and a reflux condenser.

Table 1 shows summary of cultivation conditions for all experiments.

2.3 Biomass production

The culture was monitored by optical density measurement at a wavelength of 550 nm using a Jenway 6300 spectrophotometer after appropriate dilution. Cell dry weight (CDW, g l⁻¹) of *D. communis* could be correlated to the OD (optical density) at 550 nm with a linear equation (1):

$$CDW = 0.45 \times OD \quad (1)$$

Biomass productivity was calculated from the variation in biomass concentration within a cultivation time using equation (2):

$$P = \frac{[X_1 - X_0]}{[t_1 - t_0]} \quad (2)$$

where X_0 is biomass concentration (g l⁻¹) on day t_0 ; X_1 is biomass concentration (g l⁻¹) on day t_1 .

Table 1. Plan of growth conditions for all groups.

Group	Experiment No.	Light intensity, $\mu\text{mol m}^{-2}\text{s}^{-1}$	Shaking, rpm	Bubbling, l l ⁻¹ m ⁻¹	CO ₂ concentration, v/v%
I	1	200	-	0.5	4
	2	200	-	0.5	8
II	4	200	160	0.5	4
	5	200	160	0.5	8
	6	200	160	0.5	16
	7	200	160	0.5	0
	8	200	160	0.5	1
III	9	100	160	-	-
	10	200	160	-	-
	11	300	160	-	-
IV	12	100	160	0.5	4
	13	200	160	0.5	4
	14	300	160	0.5	4

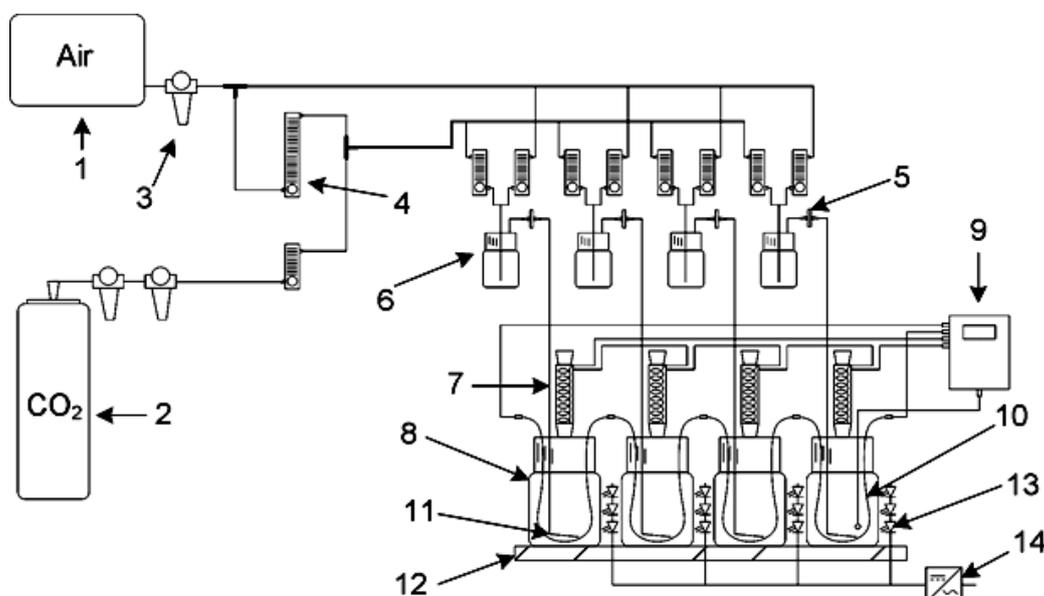


Figure 1. Schematic diagram of the experiment setup: 1 – air pump; 2 – CO₂ cylinder; 3 – reductor; 4 – gas flow meters; 5 – filter; 6 – pure water; 7 – condenser; 8 – microalgae cultures; 9 – thermostat; 10 – thermostat tubes; 11 – gas sparger; 12 – orbital shaker; 13 – LED; 14 – DC/AC adapter.

3 Results and discussion

In literature there is mentioned that mixing, light intensity and CO₂ concentration are parameters that may essentially influence the growth of algae (Sforza et al., 2012). There are different ways how the culture cultivated in flasks can be mixed. For example, it can be mixed manually – hand shaken – one or more times a day (Makarevičienė et al., 2011), it can be mixed using a shaker (Han et al., 2012) and also by means of aeration (Barsanti and Gualtieri, 2006). Information in literature on optimal light intensity and CO₂ concentration for successful microalgae growth differs (Fang et al., 2013; Samori et al., 2013; Soletto et al., 2008).

3.1 Group I – Cultivation with aeration

In order to determine the influence of mixing on microalgae growth, in Group I *D. communis* were cultivated without shaking but with aeration using two different CO₂ concentrations – 4 and 8 v/v% (gas supply was 0.5 l l⁻¹m⁻¹). The effect of these CO₂ concentrations on biomass growth is illustrated in Figure 2. As shown in Figure 2, the growth profiles of *D. communis* were very similar for CO₂ concentrations of 4 and 8 v/v%. Maximal biomass productivity for 4% CO₂ was 0.47 g d⁻¹, while for 8 v/v% CO₂ the productivity was 0.43 g d⁻¹.

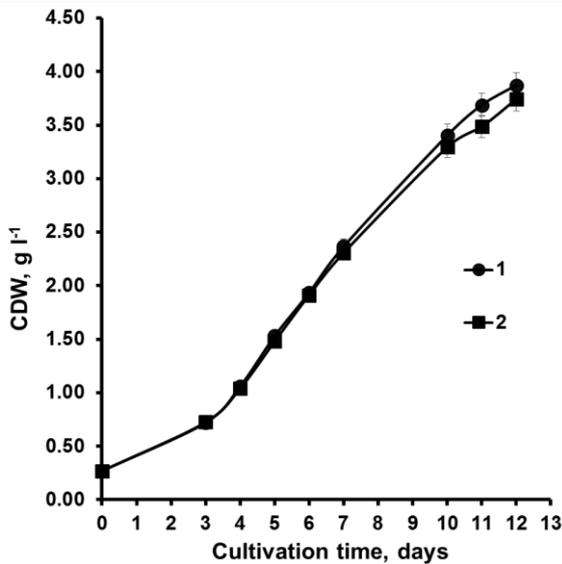


Figure 2. Effect of 4 v/v% CO₂ (1) and 8 v/v% CO₂ (2) on algae biomass growth when mixing provided only by gas bubbling.

On day 12, the accumulated DCW was 3.9 (4 v/v%) and 3.7 g l⁻¹ (8 v/v%) thus showing that even though measurements at both concentrations almost perfectly match each other, still it is CO₂ concentration of 4 v/v% that gives better results.

3.2 Group II – Cultivation with aeration and shaking

Unlike Group I, the experiments of Group II were performed using gas bubbling with three different CO₂ concentrations – 4, 8, and 16 v/v% – and a shaker, which was set up for 160 rpm providing sufficient mixing. Figure 3 illustrates the effect of CO₂ concentration on *D. communis* biomass growth.

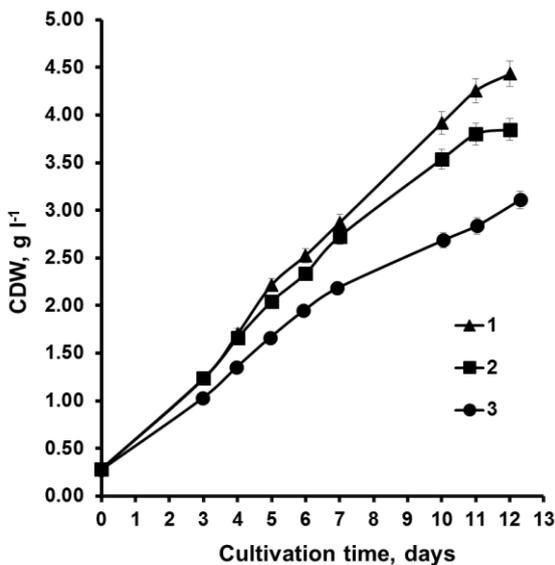


Figure 3. Effect of 4 v/v% (1), 8 v/v% (2) and 16 v/v% (3) of CO₂ on algae biomass growth when mixing provided by shaker and gas bubbling.

At the beginning of the experiment, there is no significant difference between results obtained using 4 and 8 v/v% of CO₂ concentration. The differences

appear after day 4. Using 4 v/v% CO₂, maximal biomass productivity of 0.51 g d⁻¹ was reached, but with 8 v/v% it was 0.42 g d⁻¹ and with 16 v/v% CO₂ 0.32 g d⁻¹. At the end of the experiment, the highest dry cell weight reached was 4.43 g l⁻¹ by using 4 v/v% of CO₂, while 8 and 16 v/v% yielded in 3.85 g l⁻¹ and 3.2 g l⁻¹, respectively.

Furthermore, in the experiments performed at 8 and 16 v/v% CO₂, the inhibition of culture growth was observed. It can be explained by low pH and osmotic stress. This inhibition shows that 8 and 16 v/v% concentrations are too high for *D. communis* cultivation under given conditions.

Another set of experiments using 0, 1 and 4 v/v% of CO₂ were performed in order to examine the influence of lower CO₂ concentration on algae biomass growth (Fig. 4).

As it is shown in Figure 4, the best biomass growth during the first 3 days was at low CO₂ concentration (0 and 1 v/v%). After day 6, the best growth was observed at 1 v/v% but on day 10 at 4 v/v%. This could be explained by CO₂ inhibition of culture growth at the beginning of the experiment, but later when the concentration of biomass increased, CO₂ limitation changed the growth. On the last day of cultivation experiment where air without CO₂ was used (day 13) the lowest results (2.55 g l⁻¹) yielded reaching maximal productivity of 0.43 g d⁻¹.

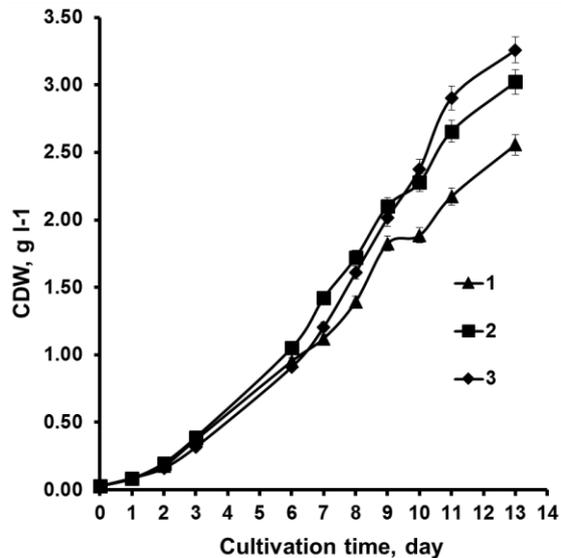


Figure 4. Effect of 0 v/v% (1), 1 v/v% (2) and 4 v/v% (3) of CO₂ on algae biomass growth when mixing provided by shaker and gas bubbling.

The highest biomass concentration (3.26 g l⁻¹) and productivity (0.53 g d⁻¹) were achieved when aeration with 4 v/v% CO₂ mixture with air was used. Thus pointing out that such a low concentration as 0 or 1 v/v% of CO₂ is not sufficient when cultivating microalgae.

By comparing Group I and II, one can conclude that the application of shaker in addition to gas sparger gives better biomass growth at the same air-CO₂ mixture. This might be because of better gas transfer from gas to liquid phase and also because good mixing prevents sedimentation of algae.

Furthermore, in both groups the best biomass growth was obtained when 4 v/v% of CO₂ mixture with air was used for aeration.

3.3 Group III – Cultivation with shaking and different light intensities

To study the effect of mixing and light intensity on algae growth, another experiment was performed where *D. communis* was cultivated with shaking and at three different light intensities (100, 200, and 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$) but without aeration. The effect of the mentioned parameters is shown in Figure 5.

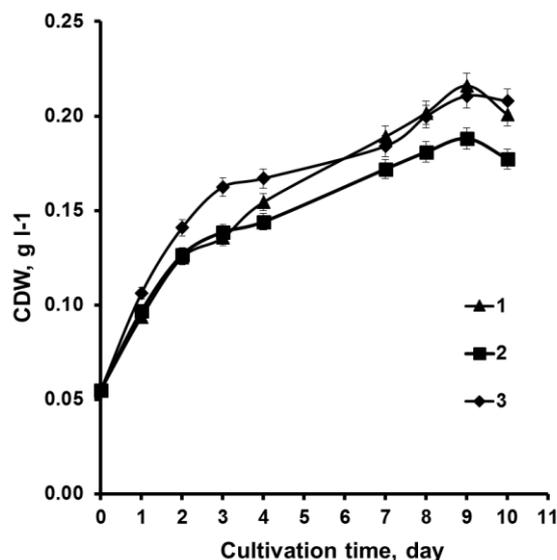


Figure 5. Effect of different light intensities – 100 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (1), 200 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (2) and 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (3) – on algae biomass growth when mixing was provided only by shaker.

Figure 5 shows that microalgae are growing very similar at all three given light intensities. The best productivity was determined at 100 $\mu\text{mol m}^{-2}\text{s}^{-1}$ 0.05 g d^{-1} . Maximal biomass yield was reached on day 9 at all light intensities but on the next day the cell lyses was observed in all bottles, which might occur due to biomass inhibition by oxygen.

Also, if one compares both experimental results of Group III with Group I and II (Fig. 2, 3 and 4), it is obvious that the results of Group I and II are better: they have higher productivity as well as maximal biomass yield, thereby showing that in order to increase biomass growth and final yield, algae cultivation should be done with aeration system enriched with CO₂, just like Samori et al. stated in their article (2013).

3.4 Group IV – Cultivation with aeration, shaking and different light intensities

Just like in Group III, the experiments of Group IV were done to determine, which light intensity gives the best effect on algal growth, their difference – the experimental set up was supplemented with an aeration system: air-CO₂ mixture of 4 v/v%.

In all bottles, the culture was supplied with air enriched with CO₂ of 4 v/v% (flow rate 0.5 l l⁻¹ min⁻¹). Figure 6 illustrates the results obtained at such conditions.

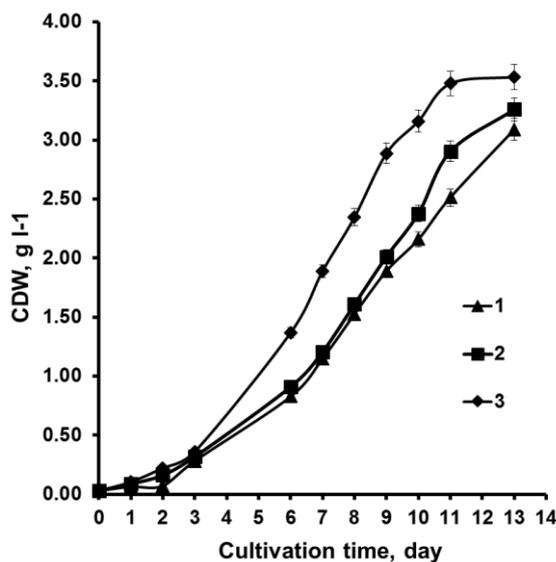


Figure 6. Effect of different light intensities – 100 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (1), 200 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (2) and 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (3) – on algae biomass growth when mixing provided by shaker and gas bubbling (4 v/v% of CO₂).

As compared to the Group III, *D. communis* is growing significantly faster. Until day 3, light intensity had a small effect on algae biomass growth but after day 3 the highest biomass growth was obtained when light intensity was the highest of the three tested – 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$. The decrease in biomass productivity at lower light intensities might be due to the light shading effect arising from the increase in cell density (Chen et al., 2011). The maximal biomass productivity and biomass yield determined at 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$ was 0.54 g d^{-1} and 3.53 g l^{-1} , respectively.

4 Conclusions

In this research, unicellular microalgae *D. communis* was used to study the effect of different cultivation conditions on microalgae biomass growth. The results showed that for successful microalgae cultivation shaking and aeration is necessary. Variation of light intensity and concentration of CO₂ can significantly affect biomass growth and yield. Best results (biomass productivity of 0.54 g d^{-1} and biomass concentration of 3.53 g l^{-1}) were obtained when mixing was provided by means of shaker and gas bubbling with CO₂ (air CO₂ mixture 4 v/v%), at light intensity 300 $\mu\text{mol m}^{-2}\text{s}^{-1}$.

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Purtymo, CO₂ koncentracijos ir šviesos intensyvumo įtaka žaliojo dumblio *Desmodesmus communis* biomasės augimui

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Dumblių augimą gali lemti daugelis veiksnių. Šiame tyrime buvo atlikti keturių skirtingų grupių eksperimentai, kuriais buvo siekiama nustatyti skirtingų maišymo sąlygų, CO₂ koncentracijos ir šviesos intensyvumo įtaką *Desmodesmus communis* augimui. CO₂ koncentracijos ribos oro–CO₂ mišinyje buvo 0–16 v/v%, šviesos intensyvumo ribos 100–300 μmol m⁻²s⁻¹. Nustatyta, kad didžiausias biomasės augimo greitis ir išeiga – atitinkamai 0.54 g d⁻¹ ir 3.53 g l⁻¹ – buvo gauti, kai maišymas buvo atliekamas purtytuvu, tiekiant 96:4 v/v% oro–CO₂ mišinį ir esant 300 μmol m⁻²s⁻¹ šviesos intensyvumui.

Raktiniai žodžiai: *dumbliai, maišymas, CO₂ koncentracija, šviesos intensyvumas, biomasės augimas.*