EREM 77/1

Journal of Environmental Research, Engineering and Management Vol. 77 / No. 1 / 2021 pp. 76–84 DOI 10.5755/j01.erem.77.1.27952 Biosorption of Cu and Zn in a Batch System via Dried Macroalgae Halimeda opuntia and Turbinaria turbinata

Received 2020/11

Accepted after revision 2021/02



ref http://dx.doi.org/10.5755/j01.erem.77.1.27952

Biosorption of Cu and Zn in a Batch System via Dried Macroalgae *Halimeda opuntia* and *Turbinaria turbinata*

Amany G. Madkour*

Marine Pollution Dept., Marine Environment Division, National Institute of Oceanography and Fisheries (NIOF), Cairo, Egypt

Mahmoud A. Dar

Marine Geology Dept., Marine Environment Division, National Institute of Oceanography and Fisheries (NIOF), Cairo, Egypt

*Corresponding author: dramanymadkour@yahoo.com

Biosorption is the most favourable technique for the treatment of heavy metals as it is fast, powerful, and low cost, it takes place in a wide range of temperatures as well as it can be used for almost all types of heavy metals. In this study, the biosorption technique adsorbs Cu^{2+} and Zn^{2+} on the dried macroalgae (*Halimeda opuntia* and *Turbinaria turbinata*) in a batch system. Experimental parameters affecting the biosorption process are initial metal ion concentrations (5, 10, 15 and 25 mg/L), pH between (4.5 and 5.2), biomass dosage (1 gm) and agitation speed 150 rpm applied at contact time (30, 60 and 120 min). The significant-high average removals of Cu^{2+} by *H.opuntia* (> 96%) were recorded in concentrations of 10, 15 and 25 ppm at 120 min and the highest average removals by *T.turbinata* (81.07%, 78.32% and 74.7%) were recorded in concentrations of 5, 10 and 15 ppm at 120 min. The lowest average removal of Cu^{2+} 89.22% was recorded by *H.opuntia* and 49.9% was recorded by *T.turbinata* in a concentration of 10 ppm at 120 min for *H.opuntia* and in a concentration of 5 ppm by at 60 min for *T.turbinata*. In conclusion, the dead biomass of marine algae can provide a promising and low-cost technique for removing heavy metal pollutants in medical industries.

Keywords: Biosorption, copper, zinc, Halimeda opuntia and Turbinaria turbinata.

Introduction

Recently, metal pollution has become a major issue all over the world, where metal ions in potable water and wastewater exceed, in many cases, the permissible sanitary levels. Many industrial activities contain dissolved heavy metals in their aqueous effluents; if these discharges are emitted without treatment, they may have adverse effects on the environment (Lu et al., 2017; Mir-Tutusaus et al., 2018). Trace amounts of heavy metals are required by living organisms, but when they exceed the permissible levels, they cause various diseases and disorders as well as deleterious ecological effects, as they are toxic and non-degradable (Mustapha & Halimoon, 2015).

Despite the importance of zinc and copper, as important minerals for living organisms, zinc at a concentration of more than 2 mg/L in wastewater causes irritation, stomach cramps, and lung disorders. Long-term exposure to high doses of copper may cause copper toxicity, which is distinguished by nausea, fever, passing out, vomiting, abdominal cramps, diarrhoea and over time causes liver damage, kidney diseases, brain damage and heart failure (Jewell, 2019). Dissolved metals cannot be removed from the natural environment, so conventional methods for their removal have been studying extensively. These methods include chemical precipitation, ion exchange/chelation, adsorption on activated carbon, and membrane processes (Sharma, 2015; Morin-Crini & Crini, 2017). However, application of these methods is often restricted because they are ineffective and costly. Consequently, biosorption has emerged as an alternative, inexpensive and effective technology (Nadeem et al., 2016).

Biosorption, the passive uptakes process to bind heavy metals on the cellular structure of biological mass, has emerged as an attractive technology due to its simplicity, high efficiency, flexibility of operation, and low cost (Espinosa et al., 2016). Other features of the biosorption technique are easy regeneration of the biosorbent and recuperation of the sorbate. The biosorption process involves absorption of heavy metals in solution on the surface of a cell wall through interaction with functional groups (i.e., carboxylate, amine, amide, imidazole, phosphate, hydroxyl, and other groups) found in cell walls of biopolymers. Many parameters affect the biosorption process efficiency such as biosorbent characteristics (i.e., permeability, surface territory), metal ion features (i.e., molecular weight, oxidation state, ionic radius), in addition to other parameters such as pH, temperature, contact time, biosorbent and sorbate concentration etc. (Escudero et al., 2019).

For years, bacteria, yeast, fungi and algae have been widely used as biosorbents for metal ions (Rahman et al., 2019). Among the biological materials, algae are the most efficient and modest biosorbents because of their slight nutrient requirement. In view of the statistical analysis on algae efficiency in the biosorption process, it has been accounted that algae retain 15.3%-84.6% percentage higher than other microbial biosorbents (Mustapha & Halimoon, 2015). The mechanism of the macroalgae biomass absorption process can be explained simply that macroalgae contain chemical groups attached to the cell wall polysaccharides and proteins such as amino, carboxyl and hydroxyl groups that react with metal ions through many reactions, i.e., complexation, coordination, microprecipitation and ion exchange (Ali Redha, 2020).

The aim of the current study was to evaluate the biosorption potential of dried macroalgal biomass for removing and recovering heavy metals ions (Cu and Zn) from a synthetic aqueous solution and to specify the effects of various factors, i.e., initial metal ion concentration and contact time at pH (4.5 to 5.2) and biosorbent dose (1 gm) for the Cu and Zn ion removal.

Material and Method

Preparation of biosorbents

One kilogram of *H.opuntia* and *T.turbinata* was collected from Red Sea coast in front of NIOF region, the collected samples then were washed with tap water followed by distilled water to remove any agglutinated materials as the epiphytes, sand or mud particles. The algae were shade dried, and then oven dried at 60°C for 48 h to remove excess water and moisture. The dried algae



biomass was cut and ground in a mechanical grinder to be a ground fine powder, subsequently sieved and the particles with an average size of 0.5 mm were used as a biosorbent (Christobel & Lipton, 2015).

Preparation of standard solutions

Cu $(No_3)_2$. $3H_2O$ and Zn $(NO_3)_2$. $6H_2O$ were used as the source of metal in solutions. Copper and zinc solutions were prepared in concentration levels at 5, 10, 15 and 25 ppm by diluting the stock solution (1000 ppm) with distilled water.

Experimental procedure

The experiment on the effects of varying initial metal ion concentrations (5, 10, 15 and 25 ppm) and contact time (30, 60 and 120 min) at pH between 4.5 to 5.2 and adsorbent dose 1gm was carried out by adding 1 g dry weight of *H.opuntia* and *T.turbinata* to a series of flasks containing 100 mL of solution of varying initial metal concentrations. The flasks were shaken at 150 rpm for 30, 60 and 120 min in the dark in a temperature-controlled shaking incubator (at 20°C) to allow the metal uptakes process (Hashim et al., 2004). After that, the samples were filtered through a GF/C filter and the filtrates were analyzed for residual metal concentration by flame atomic absorption spectrophotometer (AAS, GBC-932). We made triplicate samples to assess the accuracy and the mean values that were used for the concentration calculations. The results were expressed in terms of percentage removal of Cu and Zn ions, as given below by the equation:

% Removal = ((Co-Ce)/Co))* 100

(1)

Where Co is the initial metal ion concentration in ppm, Ce is the concentration levels of Cu and Zn ions after adsorption (ppm). (Mamatha et al., 2012).

Statistical analysis

Statistical analysis was carried out by using Minitab (version 19) to indicate a significance of treatment. Oneway analyses of variance (ANOVA) followed by Fisher's grouping test were used to indicate a significant difference between copper and zinc adsorption at different concentrations and contact times.

Results and Discussion

SEM analyses

The change in morphology of using macroalgae species after the biosorption process was predicted by using SEM spectra at 15.0 kV x 2000 resolutions for *H.opuntia* and 15.0 kV x 500 resolutions for *T.turbinata* and was shown in Plates 1 and 2. Plate (1A) and (2A) showed the raw form of using *H.opuntia* and *T.turbinata* before the biosorption process; meanwhile, plates 1 (B, C) and 2 (B, C), it showed the same species after biosorption of

Plate 1. SEM spectrum of H.opuntia before (1A) and after adsorption of Cu (1B) and Zn (1C) at 150 KV x100 and 150KV x2000





Plate 2. SEM spectrum of T.turbinata before (2A) and after adsorption of Cu (2B) and Zn (2C) at 150 KV x100 and 150KV x500

Cu and Zn ions. These plates illustrated that the pores, surfaces, small channels and hollow cylindrical shaped pores were covered and filled with Cu and Zn ions. This is evidence of the change in the initial structure of *H.opuntia* and *T.turbinata* after the biosorption process.

Effect of initial ion concentration

Ashraf et al. (2011) found that the metal biosorption capacity increases when the concentration of initial metal increases until a certain concentration. Initial metal concentration has a similar effect as the contact time. When low metal concentration is used, most of the binding sites of the biosorbent may not be occupied, so the true maximum biosorption capacity of the biosorbent would not unveil. With increasing metal concentration to the extent that the binding sites of biosorbent become fully saturated with the metal, the increase in metal concentration will not be effective. Initial ion concentration plays an important role in the biosorption process. H.opuntia was recording Cu²⁺ removal efficiency between 89.22% in 25 ppm concentration and 96.80% in 15 ppm concentration, while T.turbinata was recording removal efficiency between 49.90% in 25 ppm concentration and 81.07% in 5 ppm concentration, respectively. A slight decrease in the biosorption efficiency was observed to occur as the concentration of Cu2+ ions increased from 10 to 25 ppm with *T.turbinata*; meanwhile, the biosorption efficiency values were close to each other at 10 and 15 ppm and slightly decreased at 25 ppm with H.opuntia (Fig. 1). The estimated results indicated that *H.opuntia* was more efficient than *T.turbinata* in the removal of Cu.

Fig. 1. The effect of initial ion concentration on Cu and Zn removal efficiency by H.opuntia and T.turbinata







Fig. 2. The effect of contact time on Cu^{+2} and Zn^{+2} removal efficiency by H.opuntia and T.turbinata

In case of Zn²⁺, *H.opuntia* was recorded to show the optimum adsorption efficiency (94.98%) in 10 ppm ion concentration. Contrary to the case of Cu²⁺, there was a prominent decrease in biosorption efficiency when the concentration of Zn ions was increased from 10 to 25 ppm. On the other hand, the maximum removal efficiency of T.turbinata (94.44%) was observed in 5 ppm concentration. It was noticed that the removal efficiencies of Zn²⁺ with *H.opuntia* and *T.turbinata* were almost equal (Table 2). It is clear that increasing Zn²⁺ concentrations from 5 to 25 ppm showed a significant decrease in the removal efficiency. These illustrations indicated that the dry marine algae of both species had the highest efficient removal property in the low Zn²⁺ concentration solutions and were more effective in Cu2+ solutions with all concentrations. This illustration is in complete agreement with Ibrahim et al.'s (2016) study, where they report that the adsorption process is more effective at low concentrations because all active sites on the algal surface are vacant. Inversely, at the higher metal levels in aqueous solutions, the binding sites of the algal surfaces become consumed and finally the biosorbent surfaces become saturated and uptakes rates decrease (Al-Qodah, 2006; Tsai & Chen, 2010; Yang et al., 2010).

Effect of contact time

80

Contact time refers to the required time for the biosorption process to carry out. The contact time was one of the vital factors affecting the biosorption process efficiency. To specify experimental conditions, increasing contact time would allow the biosorbent material to detect the maximum biosorption amplitude. As the biosorbent reaches its maximum biosorption capacity, the binding sites become fully saturated and an increase in the contact time becomes worthless. Fig. 2 summarizes the biosorption efficiency of Cu and Zn ions by H.opuntia and *T.turbinata* as a function of contact time at different concentration levels. The adsorption behaviour shows variation among the studied metal ions. The rate of Cu ion removal was fast, as 89.2% to 94.30% and 49.9% to 73.00% of the metal was removed at 30 min within H.opuntia and T.turbinata, respectively. During the initial stage, a larger surface area of algae is available; therefore, the adsorption guickly occurs at the active biding sites (El-Moselhy et al., 2017). It was noticed that within 60 min contact time, removal efficiency decreased when T.turbinata was used and raised again at 120 min. In case of zinc, within 30 min, 85.8% to 92.59% and 88.05% to 90.96% were removed by *H.opuntia* and *T.turbinata*, respectively. Within H.opuntia concentrations of 15 and 25 ppm, the removal efficiency increased gradually with time until 120 min. Within T.turbinata, concentration of 5, 10, 15, and 25 ppm, the removal efficiency increased within an increase of time from 30 to 60 min, then a slight decrease occured with 5 and 15 mg/L at 120 min. Percentages of Zn²⁺ and copper removal at the definite conditions of different types of macroalgae used in previous studies are listed in Table 1.

Seaweed	Туре	Treatment conditions	Heavy Metals	Adsorption capacity	References
Halimeda opuntia	Green seaweed	Cu ²⁺ t =120 min T = 20°C Biomass = 1 g pH = 4.5–5.2 rpm = 150 lons concentration = 10, 15 and 25 mg/L Zn ²⁺ t =120 min T = 20°C Biomass = 1 g pH = 4.5–5.2 rpm = 150 lons concentration = 10 mg/L	Cu ²⁺ and Zn ²⁺	Cu ²⁺ > 96% Zn ²⁺ > 94%	Present study
Turbinaria turbinata	Brown seaweed	Cu ²⁺ t = 120m T = 20°C Biomass = 1 g pH = 4.5–5.2 rpm = 150 lons concentration =5 and 10 mg/L Zn ²⁺ t = 60 min T = 20°C Biomass = 1 g pH = 4.5–5.2 rpm = 150 lons concentration = 5 mg/L	Cu2+ and Zn2+	Cu²+ (74.7−81.07%) Zn²+ > 94%	Present study
Sargassum sp.	Brown seaweed	t = 60 min T = 25°C Biomass = 1 g pH = 3 rpm = 200 Ions concentration = 5 mg/L	Zn ^{2*}	Zn ²⁺ = 90.3% (acid treated)	(Mahmood et al., 2017)
Sargassum sp.	Brown seaweed	t = 6 h $T = 30^{\circ}C$ Biomass = 0.1 g pH = 5 rpm = 150 Ion concentration 0–7 mmol/L	Cu ²⁺	Cu ²⁺ = 1.483 mmol/g	(Barquilha et al., 2017)
Sargassum filipendula	Brown seaweed	t = 24 h T = 25°C Biomass = 2 g/L pH = 3.5 rpm = 180 Ion concentration = 1 mmol/L	Cu^{2*} and Zn^{2*}	Cu ²⁺ = 69.05% Zn ²⁺ = 44.21%	(Cardoso et al., 2017)

Table 1. Comparison between the obtained biosorption capacity of Cu²⁺ and Zn²⁺ within Halimeda opuntia and Turbinaria turbinata and some pervious studied macroalgal species

81

Seaweed	Туре	Treatment conditions	Heavy Metals	Adsorption capacity	References
Osmundea pinnatifida	Red seaweed	t = 180 and 60 min T = 20 to 23°C Biomass = 20 g/L pH = 5 rpm = 500 Ion concentration=100 and 50 mg/L	Cu ²⁺	Cu ²⁺ = 50.89% to 71.64%	(El Hassouni et al., 2014)
Fucus vesiculou	Brown algae	t = 2h $T = 25^{\circ}C$ Biomass = 0.25 g/L pH = 5 Ion concentration=10–150 mg/L		0.97 mg/g	(Brinza et al., 2007)
Gracilaria fisheri	Red algae	t = 30m $T = 22^{\circ}C$ Biomass = 2 g/L pH 4 rpm = 100 Ion concentration = 1 mM	Cu ²⁺	90% (treated with NaoH) 80% (native)	(Chaisuksant, 2003)
Ulva sp	Green algae	t = 2 h T = 22°C Biomass = 8 g/L pH = 5.5 Ion concentration = 10 to 220mg/L	Zn²+	29.63 mg/g	(Badescu et al., 2017)
Ulva fasciata and Sargassum sp.	Green algae Brown algae	t = 30 m T =22°C Biomass = 0.1 to 5 g/L pH 5.5 rpm = 150 Ion concentration = 10 to 220 mg/L	Cu ²⁺	73.5 mg/g for <i>U. fasciata</i> and 72.5 mg/g for <i>Sargassum sp</i>	(Karthikeyan et al., 2007)

Table 1 (continued)

Data analysis

The ANOVA result indicates highly significant differences between two tested seaweeds in the removal of copper and zinc at different concentrations (P = 0.001, Tables 2, 3). In addition, Fisher's grouping test indicates a significant difference between copper and zinc at different concentrations and times. The highest removal of copper was 96.8%, 96.27%, and 96.19% at concentrations of 15, 25, and 10 ppm, respectively, by *H.opuntia* at 120 min, while the lowest removal of copper was 49.9%

Table 2. ANOVA and Fisher	's grouping test of remova	ıl (%) of Cu at different co	oncentration (ppm) and time (min)

	ANOVA		Time (min)	Fisher's test for different concentrations (ppm) grouping			
	F	Р		5	10	15	25
	373.1	0.001	30	$93.34\pm0.25^{\text{B}}$	$94.30\pm0.13^{\rm A}$	93.40 ± 1.49^{B}	89.22 ± 0.22 ^c
H.opuntia	140.3	0.001	60	92.14 ± 0.247^{B}	95.26 ± 0.24^{A}	95.55 ± 0.322 ^A	92.21 ± 0.29 ^B
	312.9	0.001	120	92.8 ± 0.16 ^c	96.19 ± 0.09^{B}	96.8 ± 0.21 ^A	96.27 ± 0.21 ^B
	437.2	0.001	30	$73 \pm 0.06^{\text{A}}$	71.2 ± 0.1 ^B	61.22 ± 011 ^c	49.9 ± 0.06 ^D
T.turbinata	191.9	0.001	60	60.22 ± 0.07^{B}	$64.52 \pm 0.08^{\text{A}}$	56.59 ± 0.04 ^c	52.6 ± 0.05 ^D
	521.3	0.001	120	81.07 ± 0.02 ^A	78.32 ± 0.05^{B}	74.7 ± 0.02 ^c	70.4 ± 0.01 ^D

	ANOVA		Time (min)	Fisher's test for different concentrations (ppm) grouping			
	F	Р	Tittle (Tillit.)	5	10	15	25
H.opuntia	902.3	0.001	30	$92.59\pm0.04^{\rm A}$	91.3 ± 0.05^{B}	$88 \pm 0.08^{\circ}$	85.8 ± 0.01 ^D
	197.8	0.001	60	$92.88\pm0.01^{\rm A}$	90.29 ± 0.03^{B}	89.7 ± 0.03 ^c	86.88 ± 0.02 ^D
	177.6	0.001	120	$92.37 \pm 0.09^{\text{A}}$	94.98 ± 0.09^{B}	93.19 ± 0.01 ^c	90.84 ± 0.04^{D}
T.turbinata	182.6	0.001	30	$88.05\pm0.08^{\rm A}$	88.89 ± 0.02^{B}	90.69 ± 0.01 ^c	90.96 ± 0.07 ^D
	805.2	0.001	60	$94.44\pm0.05^{\rm A}$	92.44 ± 0.1^{B}	92.16 ± 0.04 ^c	91.34 ± 0.09 ^D
	628.2	0.001	120	$91.83\pm0.05^{\scriptscriptstyle B}$	$93.65\pm0.05^{\rm A}$	$90.09\pm0.07^{\text{D}}$	91.45 ± 0.11 ^c

Table 3. ANOVA and Fisher's grouping test of removal (%) of Zn at different concentration (ppm) and time (min)

at concentrations 25 ppm by *Turbinaria turbinata* at 30 min. As for zinc, the highest removal was 94.9% at concentrations of 10 ppm by *H.opuntia* at 120 min, followed by 94.4% at a concentration of 5 ppm by *T.turbinata* at 60 min, while the lowest removal was 85.8% at a concentration 25 ppm by *H. opuntia* at 30 min.

Conclusion

Based on the obtained results, it can be concluded that the studied species (*H.opuntia* and *T.turbinata*) have

References

Ali Redha A. (2020) Removal of heavy metals from aqueous media by biosorption. Arab Journal of basic and applied sciences 27(1): 183-193. https://doi.org/10.1080/25765299.2020.1756177

Al-Qodah Z. (2006) Biosorption of heavy metal ions from aqueous solutions by activated sludge. Desalination 196(1-3): 164-176. https://doi.org/10.1016/j.desal.2005.12.012

Ashraf M.A., Abdul Wajid, Mahmood K., Maah M.J. and Yusoof I. (2011) Low cost biosorbent banana peel (Musa sapientum) for the removal of heavy metals. Scientific Research and Essays, 6(19): 4055-4064. https://doi.org/10.5897/SRE11.303

Badescu I.S., Bulgariu D. and Bulgariu L. (2017) Alternative utilization of algal biomass (Ulva sp.) loaded with Zn (II) ions for improving of soil quality. Journal of Applied Phycology, 29: 1069-1079. https://doi.org/10.1007/s10811-016-0997-y

Barquilha C.E.R., Cossich E.S., Tavares C.R.G. and Silva E.A. (2017) Biosorption of nickel (II) and copper (II) ions in batch and fixed-bed columns by free and immobilized marine algae Sargassum sp. Journal of Cleaner Production, 150: 58-64. https:// doi.org/10.1016/j.jclepro.2017.02.199

Brinza L., Dring M.J. and Gavrilescu M. (2007) Marine micro and macro algal species as biosorbents for heavy metals. Environ-

high abilities for copper and zinc removal at low concentrations. Both species are able to remove significant amounts of copper and zinc, hence supporting their potential application in the treatment of wastewater polluted by the metals. *H.opuntia* has higher efficiency for copper removal in comparison to *T.turbinata*, where more than 96% of copper is removed at 120 min within *H.opuntia*, while *T.turbinata* removes 81% of Cu ions at 120 min. In case of zinc, the twoused macroalgae have close competencies; they remove more than 85% within 30 min.

mental Engineering and Management Journal, 6(3): 237-251. https://doi.org/10.30638/eemj.2007.029

Cardoso S.L., Costa C.S.D., Nishikawa E., da Silva M.G.C. and Vieira M.G.A. (2017) Biosorption of toxic metals using the alginate extraction residue from the brown algae Sargassum filipendula as a natural ion-exchanger. Journal of Cleaner Production, 165: 491-499. https://doi.org/10.1016/j.jclepro.2017.07.114

Chaisuksant Y. (2003) Biosorption of cadmium (II) and copper (II) by pretreated biomass of marine alga Gracilaria fisheri. Environmental Technology, 24(12): 1501-1508. https://doi.org/10.1080/0959330309385695

Christobel J. and Lipton A.P. (2015) Evaluation of macroalgal biomass for removal of heavy metal Arsenic (As) from aqueous solution. International Journal of Application or Innovation in Engineering & Management, 4(5): 94-104. ISSN: 2319-4847. https:// ijaiem.org/pabstract_Share.php?pid=IJAIEM-2015-04-13-35

El Hassouni H., Abdellaoui D., El Hani S. and Bengueddour, R. (2014) Biosorption of cadmium (II) and copper (II) from aqueous solution using red alga (Osmundea pinnatifida) biomass. Journal of Materials and Environmental Science, 5(4): 967-974.



Available on: https://www.jmaterenvironsci.com/Document/ vol5/vol5_N4/119-JMES-612-2014-elhassouni.pdf

El-Moselhy K.M., Abdel Azzem M., Amer A. and Al-Prol A.E. (2017) Adsorption of Cu(II) and Cd(II) from aqueous solution by using rice husk adsorbent. Physical Chemistry Indian Journal 12(2): 109-122. https://www.tsijournals.com/articles/adsorption-of-cuii-and-cdii-from-aqueous-solution-by-using-rice-husk-adsorbent-13239.html

Escudero L.B., Quintas P.Y., Wuilloud R.G. and Dotto G.L. (2019) Recent advances on elemental biosorption. Environmental Chemistry Letters 17: 409-427 (2019). https://doi.org/10.1007/ s10311-018-0816-6

Espinosa-Ortiz E.J., Rene E.R., Pakshirajan K., van Hullebusch E.D. and Lens P.N.L. (2016) Fungal pelleted reactors in wastewater treatment: Applications and perspectives. Chemical Engineering Journal 283: 553-571. https://doi.org/10.1016/j. cej.2015.07.068

Hashim M.A. and Chu K.H. (2004) Biosorption of cadmium by brown, green and red seaweeds. Chemical Engineering Journal 97(2-3): 249-255. https://doi.org/10.1016/S1385-8947(03)00216-X

Ibrahim W.M., Hassan A.F. and Azab Y.A. (2016) Biosorption of toxic heavy metals from aqueous solution by Ulva lactuca activated carbon. Egyptian Journal of Basic and applied science 7(3): 241-249. https://doi.org/10.1016/j.ejbas.2016.07.005

Jewell T. (2019) What to Know About Copper Toxicity. healthline. com on March 8, 2019. https://www.healthline.com/health/copper-toxicity

Karthikeyan S., Balasubramanian R. and Iyer C.S.P. (2007) Evaluation of the marine algae Ulva fasciata and Sargassum sp. for the biosorption of Cu (II) from aqueous solutions. Bioresource Technology, 98(2): 452-455. https://doi.org/10.1016/j. biortech.2006.01.010

Lu T., Zhang Q. and Yao S. (2017) Efficient decolorization of dye-containing wastewater using mycelial pellets formed of marine-derived Aspergillus niger. Chinese Journal of Chemical Engineering 25(3): 330-337. https://doi.org/10.1016/j.cjche.2016.08.010

Mahmood Z., Zahra S., Iqbal M., Raza M.A. and Nasir S. (2017) Comparative study of natural and modified biomass of Sargassum sp. for removal of Cd2+ and Zn2+ from wastewater. Applied Water Science, 7: 3469-3481. https://doi.org/10.1007/ s13201-017-0624-3 Mamatha J., Suresh V., Vedamurthy A. and Shilpi B. (2012) Production of α-Amylase from Aspergillus flavus under solid state fermentation with optimum conditions. International Research Journal of Pharmacy 3(8): 135-140.Available at: http://www. irjponline.com/admin/php/uploads/1300_pdf.pdf

Mir-Tutusaus J.A., Baccar R., Caminal G. and Sarrà M. (2018) Can white-rot fungi be a real wastewater treatment alternative for organic micropollutants removal? A review. Water Research 138(1): 137-151. https://doi.org/10.1016/j.watres.2018.02.056

Morin-Crini N., Crini G. and Roy L. (2017) Eaux industrielles contaminées: Réglementation, parameters chimiques et biologiques & procédés d'épuration innovants, Presses universitaires de franche-comté (pufc), pp. 512. ISBN: 978-2-84867-589-3. Available at: https://pufc.univ-fcomte.fr/eaux-industrielles-contaminees.html https://doi.org/10.4000/books.pufc.10857

Mustapha M.U. and Halimoon N. (2015) Microorganisms and Biosorption of Heavy Metals in the Environment: A Review Paper. Journal of Microbial & Biochemical Technology 7: 253-256. https://doi.org/10.4172/1948-5948.1000219

Nadeem R., Manzoor Q., Iqbal M. and Nisar J. (2016) Biosorption of Pb(II) onto immobilized and native Mangiferaindica waste biomass. Journal of Industrial and Engineering Chemistry 35: 185-194. https://doi.org/10.1016/j.jiec.2015.12.030

Rahman Z., Thomas L., Singh V.P. (2019) Biosorption of heavy metals by a lead (Pb) resistant bacterium, Staphylococcus hominis strain AMB-2. Journal of Basic Microbiology 59(5): 477-486. https://doi.org/10.1002/jobm.201900024

Sharma S.K. (2015) Green Chemistry for Dyes Removal from Wastewater: Research Trends and Applications. Scrivener Publishing LLC Wiley, Beverley, pp 496. https://doi.org/10.1002/9781118721001

Tsai W.T. and Chen H.R. (2010) Removal of malachite green from aqueous solution using low-cost chlorella based biomass. Journal of Hazardous Materials 175(1-3): 844-849. https://doi. org/10.1016/j.jhazmat.2009.10.087

Yang C.P., Wang J.Q., Lei M., Xie G.X., Zeng G.M. and Luo, S.L. (2010) Biosorption of zinc(II) from aqueous solution by dried activated sludge. Journal of Environmental Sciences 22(5): 675-680. https://doi.org/10.1016/S1001-0742(09)60162-5



This article is an Open Access article distributed under the terms and conditions of the Creative Commons Attribution 4.0 (CC BY 4.0) License (http://creativecommons.org/licenses/by/4.0/).